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The microorganism has been deposited with the American Type Culture Collection under number ATCC 53552.

(54)

Enzymatic peracid bleaching system.

(57)

An enzymatic perhydrolysis system, useful for bleaching, has a novel enzyme, a substrate, and a source of hydrogen peroxide, and provides *in situ* formation of peracid in aqueous solution. The substrate is selected for enzyme catalyzed reaction, and preferably is an acylglycerol with two or three fatty acid chains. The enzyme is hydrolytically and perhydrolytically active even in the presence of anionic surfactants, has lipase activity, and is isolatable from *Pseudomonas putida* ATCC 53552.

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Descripti n

ENZYMATIC PERACID BLEACHING SYSTEM

This invention relates to an enzymatic peracid bleaching system, more particularly, it relates to an enzymatic perhydrolysis system and to the use of such a system in aqueous solution for achieving enhanced bleaching.

Various bleaches have long been employed in numerous cleaning applications, including the washing and prewashing of fabrics as well as in other applications, such as hard surface cleaning. In these applications, the bleaching agent oxidizes various stains or soils on fabrics, textiles and hard surfaces.

Peroxygen bleaching compounds such as hydrogen peroxide, sodium percarbonate and sodium perborate have been found useful in dry bleach formulations because of their oxidizing power.

It has also been found that addition of certain organic compounds, including activators such as tetracetyl ethylene diamine, to perborate bleaches can improve bleaching performance because of in situ formation of peracids.

Cleaning compositions for fabrics, textiles and other materials including hard surfaces have also been developed which employ various enzymes for removing certain stains or soils. For example, protease enzymes have been found useful for hydrolyzing protein-based stains particularly in the cleaning of fabrics. Amylase enzymes have been found useful against carbohydrate-based stains resulting for example from foods. Lipase enzymes have also been found useful for hydrolyzing fat-based stains in a prewash or resoak mode.

In connection with the use of enzymes in cleaning or detergent compositions, European Patent Application, Publication No. 0 130 064, applied for by Novo Industry A/S, related to improvements in enzymatic additives for use with detergents in washing applications. That publication discussed the use of lipase enzymes for achieving substantially improved lipolytic cleaning efficiency, over a broad range of wash temperatures including relatively low temperatures below 60°C. This reference further disclosed the use of enzymes, including lipases, for direct interaction with stains or soils as a means of at least partially dissolving or loosening such fat-based stains.

U.S. Patent 3,974,082, issued August 10, 1976 to Weyn, disclosed a bleaching composition and method of use in which an alkyl ester was combined with an esterase or lipase enzyme in an aqueous medium and which was contended to liberate an acyl moiety from the ester. The Weyn patent further contended that the use of this combination together with a percompound would lead to in situ formation of peracid.

Accordingly, there has been found to remain a need for improved bleaching or activated oxidant systems capable of enhanced performance in aqueous solution at low temperature wash conditions while still maintaining high temperature performance.

The present invention provides an activated oxidant system for achieving enzymatic perhydrolysis of a substrate in the presence of a source of hydrogen peroxide to produce a peracid. A novel enzyme of the invention acts catalytically to enhance the reaction of substrates resulting the in situ formation of peracids. This novel enzyme has excellent perhydrolysis characteristics and good reactivity for triglyceride substrates, even in the presence of anionic surfactants. Thus, the enzymatic perhydrolysis system of the present invention is compatible with commercially available detergents utilizing anionic surfactants.

The novel enzyme is isolatable from Pseudomonas putida ATCC 53552, and has the following amino acid sequence:

0 268 456

1	ala	pro	leu	pro	asp	thr	pro	gly	ala	10	pro	phe	pro	
								20						5
	ala	val	ala	asn	phe	asp	arg	ser	gly		pro	tyr	thr	
	thr	ser	ser	gln	ser	30	gly	pro	ser		cys	arg	ile	
														10
	tyr	arg	pro	40	arg	asp	leu	gly	gln	gly	gly	val	arg	
	his	50	pro	val	ile	leu	trp	gly	asn	gly	thr	gly	60	ala
														15
	gly	pro	ser	thr	tyr	ala	gly	leu	leu	70	ser	his	trp	
	ala	ser	his	gly	phe	val	val	80	ala	ala	ala	glu	thr	
														20
	ser	asn	ala	gly	thr	90	gly	arg	glu	met	leu	ala	cys	
	leu	asp	tyr	100	leu	val	arg	glu	asn	asp	thr	pro	tyr	
														25
	gly	110	thr	tyr	ser	gly	lys	leu	asn	thr	gly	arg	120	val
	gly	thr	ser	gly	his	ser	gln	gly	gly	130	gly	gly	ser	
														30
	ile	met	ala	gly	gln	asp	thr	140	arg	val	arg	thr	thr	
	ala	pro	ile	gln	pro	150	tyr	thr	leu	gly	leu	gly	his	
														35
	asp	ser	ala	160	ser	gln	arg	arg	gln	gln	gly	pro	met	
	phe	170	leu	met	ser	gly	gly	gly	asp	thr	ile	ala	180	phe
														40
	pro	tyr	leu	asn	ala	gln	pro	val	tyr	190	arg	arg	ala	
	asn	val	pro	val	phe	trp	gly	200	glu	arg	arg	tyr	val	
														45
	ser	his	phe	glu	pro	210	val	gly	ser	gly	gly	ala	tyr	
	arg	gly	pro	220	ser	thr	ala	trp	phe	arg	phe	gln	leu	
														50
	met	230	asp	asp	gln	asp	ala	arg	ala	thr	phe	tyr	240	gly
	ala	gln	cys	ser	leu	cys	thr	ser	leu	250	leu	trp		
														55
	ser	val	gly	arg	arg	gly	leu							
														60

The substrate of the activated oxidant system is selected for enzyme catalyzed reaction, in the presence of a source of hydrogen peroxide, to form peracid. Various triglycerides are particularly suitable for forming the

substrate. Particularly preferred substrates are trioctanoin and tridecanoin.

The oxidant system of the invention includes a source of hydrogen peroxide which will combine with the substrate, when activated by the enzyme, to produce in situ an organic peracid. For United States laundry conditions, a particularly preferred organic peracid so generated is peroctanoic acid.

The enzymatic perhydrolysis system of the invention provides a number of advantages, including the employment of a relatively inexpensive substrate together with a small amount of an enzyme, for producing the resulting peracid. The preferred triglyceride substrates provide the ability to yield a higher concentration of peracid then provided by equivalent concentrations of a simple ester substrate. The enzymatic perhydrolysis system of the invention has been found to be very effective for producing peracid in low temperature wash solution.

In the drawings:

Figure 1 is a map of the 4.3 kb EcoRI fragment of a plasmid designated pSNE4. The region crosshatched represents signal peptide codons (codons -22 to +1) and the stippled region indicates the coding region (codons +1 to +258) for the mature polypeptide designated Lipase 1. The ATG initiation codon and TAA stop codon are also marked; and

Figure 2 illustrates an *E. coli* expression vector for *Pseudomonas* Lipase 1 gene. The stippled region indicates the coding region for the lipase signal sequence of 22 amino acids. The crosshatched region indicates the coding region for the mature lipase protein. Transcription starts at the ATG initiation codon and proceeds in the direction indicated by the arrow to the TAA stop codon. The dark regions on either side indicate the 5'- and 3'-untranslated regions.

The enzymatic perhydrolysis system of the present invention essentially comprises a novel enzyme as defined below, a substrate, and a source of hydrogen peroxide. Accordingly, the invention is based upon peracid or perhydrolysis chemistry. Although a detailed discussion of basic peracid and perhydrolysis chemistry is not believed necessary for an understanding of the invention by those skilled in the art, the article by Sheldon N. Lewis, supra, is incorporated herein as though set out in its entirety to assist in understanding of the invention.

In addition to the essential components of the enzymatic perhydrolysis system including a novel enzyme, a substrate and a hydrogen peroxide source, the perhydrolysis system of the invention also preferably includes one or more emulsifying agents selected for maintaining the substrate in suspension, or solubilizing, the substrate when in aqueous solution and for promoting interaction of the substrate and enzyme in the presence of hydrogen peroxide from the hydrogen peroxide source. Use of one or more emulsifying agents of this type is particularly contemplated so that the emulsifying agent can assist in forming a liquid phase interface at which the enzyme can better interact with a glyceride substrate. The perhydrolysis system may also preferably include various buffering agents, stabilizers and other adjuncts described in greater detail below.

In order to ensure proper understanding and interpretation of the invention, including the summary and preferred embodiments as well as the claims, some definitions are set forth below to clarify the use of terms employed herein. The defined terms include the following.

"Perhydrolysis" is defined as the reaction of a selected substrate with peroxide to form a peracid and water.

In the preferred perhydrolysis reactions yielding a peracid, both the peroxide starting material and the peracid product are oxidants. Traditionally, inorganic peroxide has been used as the oxidant, for example, in dry laundry bleaches. The peracid product is usually the desired oxidant for laundry bleaches according to the present invention, since the oxidative ability of the peracid product makes it an effective stain removal agent for laundry bleaches, yet the peracid product as oxidant remains sufficiently mild to assure only minimal reaction with dyes and other adjuncts used in laundry bleach products. However, it is within the scope of the present invention that the enzymatic perhydrolysis system be combined with a chemical perhydrolysis system.

"Chemical perhydrolysis" generally includes those perhydrolysis reactions in which an activator or peracid precursor is combined with a source of hydrogen peroxide. A type of peracid precursor for chemical perhydrolysis is disclosed in copending Application 87 308 555.9. This specification describes sulfonated phenyl esters of dicarboxylic acids which are water soluble and give peroxyacids upon dissolution in water with a source of peroxide by means of in situ chemical perhydrolysis.

"Enzymatic perhydrolysis" is defined as a perhydrolysis reaction which is assisted or catalyzed by an enzyme generally classified as a hydrolase, and more specifically identified below.

Characteristics and preferred examples of the three essential components of the enzymatic perhydrolysis system are first discussed below, followed by a brief discussion of other adjuncts which may be used with the perhydrolysis system and then a number of examples illustrating the enzymatic perhydrolysis system of the invention.

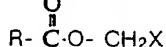
As noted above, the substrate of the enzymatic perhydrolysis system is selected for enzyme catalyzed reaction, in the presence of a source of hydrogen peroxide, to form peracid. As will be discussed in greater detail below, certain substrates are normally present as solids and particularly lend themselves to use in dry formulations including the substrate, enzyme and peroxide source. In such products, it is important that the dry formulation exhibit prolonged shelf life with the enzyme catalyzed reaction not taking place until the formulation is added to an aqueous solution.

For use in a laundry detergent formulation, for example, the substrate may also include surfactant characteristics so that in situ formation of the peracid occurs at or near the surface of the fabric to be cleaned.

This assures greater effectiveness of the oxidant responsible for bleaching action.

The substrate of the enzymatic perhydrolysis system may be chosen from a variety of different esters (RCOOR'), as the novel enzyme has esterase activity, may be a lipid, as the novel enzyme also has lipase activity, or may be a combination of both (e.g. fatty acid ester lipids). It has been found, in accordance with the present invention, that various fatty acid or glyceride type materials are particularly suitable for forming the substrate of the present enzymatic perhydrolysis system.

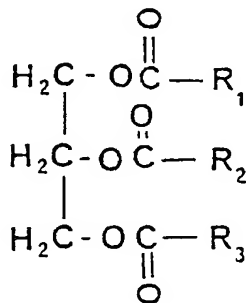
Preferably, the substrate of the present invention is a functionalized ester having the structure



wherein R is a substituent having at least one carbon atom, more preferably where R is a straight chain or branched chain alkyl optionally substituted with one or more functional groups or heteroatoms such as a phenol group, a halo group or atom, etc. and X is a functional moiety. The substrate is capable of enzymatic hydrolysis as defined above and preferably is ordinarily incapable of substantial chemical perhydrolysis. More preferably, the functional moiety comprises a functionalized polyol or polyether. More broadly, the functional moiety includes at least one carbon atom and at least one functional group.

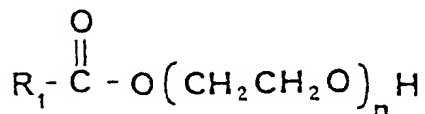
Even more preferably, the substrate of the invention is selected from the group consisting essentially of:

(i) glycerides having the structure



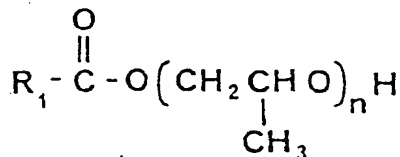
wherein $\text{R}_1 = \text{C}_1\text{-C}_{12}$, $\text{R}_2 = \text{C}_1\text{-C}_{12}$ or H and $\text{R}_3 = \text{C}_1\text{-C}_{12}$ or H;

(ii) an ethylene glycol derivative or ethoxylated ester having the structure



wherein $n = 1\text{-}10$ and R_1 is defined above; and

(iii) a propylene glycol derivative or propoxylated ester having the structure



wherein R_1 and n are defined above.

Within the preferred structures referred to immediately above, R_1 is more preferably $\text{C}_8\text{-C}_{10}$ and most preferably $\text{C}_7\text{-C}_9$, R_2 is more preferably $\text{C}_6\text{-C}_{10}$ or H and most preferably $\text{C}_7\text{-C}_9$ or H and R_3 is more preferably $\text{C}_5\text{-C}_{10}$ or H and most preferably $\text{C}_7\text{-C}_9$ or H. In structure (i) above, R_1 , R_2 and R_3 can be different chain lengths, and mixtures of such different glycerides are suitable in the present invention.

Glycerides undergo hydrolysis when boiled with acids or bases or by the action of lipases. The use of glycerides (that is, acylglycerols), especially diglycerides (diacylglycerols) and triglycerides (triacylglycerols), is particularly preferred within the enzymatic perhydrolysis system of the present invention since each triglyceride molecule is capable of yielding up to three fatty acid or peracid molecules on an equivalence basis. Thus, the use of such a glyceride may be particularly effective in achieving maximum oxidizing power in the presence of a peroxide source and enzyme as discussed in greater detail below.

Broadly, the glyceride substrate is characterized by fatty acid chains including from about 1 to about 18 carbon atoms. Lower molecular weight glycerides derived from such products as acetic acid naturally occur as liquids. Thus, additional processing steps may be necessary in order to include such a substrate in a dry formulation such as laundry detergent. However, the lower molecular weight glyceride products may also tend to be more effective in higher temperature cleaning applications.

High molecular weight glyceride substrates, such as stearic acid characterized by a chain of 17 carbon atoms, normally appear as solids and thus may facilitate their inclusion in a dry detergent formulation, for example. However, such high molecular weight fatty acid chains may not produce maximum oxidizing power in accordance with the present invention.

The most preferred form of substrate for use within the enzymatic perhydrolysis system of the invention has been found to be either trioctanoin or tridecanoin characterized respectively by fatty acid chains (including the carbonyl carbon) of 8 and 10 carbon atoms.

These two triglycerides also tend to be present as solids and thus lend themselves to inclusion in a dry formulation as discussed above. At the same time, trioctanoin and tridecanoin tend to exhibit surfactant characteristics within aqueous solution lending themselves to in situ formation of peracid as discussed above.

All of the substrates discussed above including triglycerides, such as the most preferred trioctanoin and tridecanoin, are relatively inexpensive and are thus also important for reducing initial cost of the enzymatic perhydrolysis system of the present invention. As will also be discussed below, the substrate and hydrogen peroxide source are the two major components of the enzymatic perhydrolysis system in that the enzyme need only be present in very small amounts, less than stoichiometric, to carry out the in situ peracid production contemplated in the aqueous solution. The enzyme thus acts in a catalytic manner in that, while it participates in the reaction, it is not consumed but regenerates itself for further reaction.

Virtually any source of peroxide is satisfactory as the oxidant source of the enzymatic perhydrolysis system of the invention. For example, the peroxide source may comprise a perborate or percarbonate such as sodium perborate or sodium percarbonate. In addition, the peroxide source may comprise or include hydrogen peroxide adducts such as urea hydrogen peroxide, etc.

Preferred sources of peroxide include sodium perborate monohydrate, sodium perborate tetrahydrate, sodium carbonate peroxyhydrate, sodium pyrophosphate peroxyhydrate, urea peroxyhydrate, sodium peroxide, and mixtures thereof. Sodium perborate monohydrate and sodium perborate tetrahydrate are particularly preferred as alkaline sources of peroxide.

The source of peroxide (that is, compounds yielding hydrogen peroxide in an aqueous solution) itself constitutes a peroxygen bleaching compound. However, the enzymatic perhydrolysis system provides improved bleaching. Accordingly, further discussion of the particular oxidant source is not believed necessary except to the extent that the source is selected to produce hydrogen peroxide also in accordance with the preceding discussion.

The enzyme exists in a hostile, oxidizing environment during use of the enzymatic perhydrolysis system due to the presence of peroxide and the desired peracid. Either peroxide or peracid could inactivate an enzyme in use, so an enzyme suitable for the invention must be sufficiently perhydrolytically active at expected ranges of hydrogen peroxide and at desired ranges of peracid.

The novel enzyme (sometimes herein referred to as "Lipase 1") is secreted by and isolatable from Pseudomonas putida. Pseudomonas is a genus of short, rod-shaped bacteria. Several strains, including P. putida, have been shown to have a limited ability to grow on a minimal media with mono-oleate polyoxyethylene ("Tween 80", available from Atlas Chemical) as carbon source. Howe et al., J. Gen. Microbiol., 92(1), pp. 234-235 (1976). A culture of a novel Pseudomonas putida strain from which the Lipase 1 enzyme may be isolated has been deposited in accordance with MPEP 608.1(P) in the permanent culture collection of the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852, and has been designated ATCC 53552.

It should be understood that the microorganism of the present invention is not limited to the Pseudomonas putida strain hereinabove described, as natural and artificial mutants of the said microorganism can be used. Further, genetic engineering techniques applicable to lipase production, such as transformation of corresponding genes of the present strain to other cells, may also be applied, and the lipase produced by these techniques and then isolated is of course included in the present invention.

For production, a lipase producing microorganism belonging to the genus Pseudomonas may be cultured in a conventional medium for enzyme isolation and purification. Liquid or solid culture can be used. Submerged aeration culture is preferable for industrial production. A conventional nutrient medium can be used.

Culturing temperature may vary depending on the desired rate of growth of the microorganisms and is preferably at 25°-35°C. Culturing time can be selected as desired, and is 15-50 hours. Culturing may be terminated when the highest concentration of lipase is present in the medium.

Lipase is accumulated in the fermentation broth and extraction of the produced enzyme from the broth can be effected as follows:

Cells and cell debris are first removed from the whole cell broth culture by microfiltration and centrifugation, followed by ultrafiltration to concentrate the lipase. Excess salts and color are then removed by dialysis or diafiltration. The crude enzyme solution can then be purified by conventional purification methods for proteins. A powder of the enzyme can be obtained by lyophilization and used in compositions of the invention.

The Lipase 1 has the following amino acid sequence:

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1	ala	pro	leu	pro	asp	thr	pro	gly	ala	10	pro	phe	pro	
	ala	val	ala	asn	phe	asp	arg	20	ser	gly	pro	tyr	thr	5
	thr	ser	ser	gln	ser	30	glu	gly	pro	ser	cys	arg	ile	
	tyr	arg	pro	40	arg	asp	leu	gly	gln	gly	gly	val	arg	10
	his	50	pro	val	ile	leu	trp	gly	asn	gly	thr	gly	60	ala
	gly	pro	ser	thr	tyr	ala		gly	leu	leu	70	ser	his	15
	ala	ser	his	gly	phe	val	val	80	ala	ala	ala	glu	thr	
	ser	asn	ala	gly	thr	90	gly	arg	glu	met	leu	ala	cys	20
	leu	asp	tyr	100	leu	val	arg	glu	asn	asp	thr	pro	tyr	
	gly	110	thr	tyr	ser	gly	lys	leu	asn	thr	gly	arg	120	val
	gly	thr	ser	gly	his	ser		gln	gly	gly	130	gly	gly	ser
	ile	met	ala	gly	gln	asp	thr	140	arg	val	arg	thr	thr	30
	ala	pro	ile	gln	pro	150	tyr	thr	leu	gly	leu	gly	his	35
	asp	ser	ala	160	ser	gln	arg	arg	gln	gln	gly	pro	met	
	phe	170	leu	met	ser	gly	gly	gly	asp	thr	ile	ala	180	phe
	pro	tyr	leu	asn	ala	gln	pro	val	tyr		190	arg	arg	ala
	asn	val	pro	val	phe	trp	gly	200	glu	arg	arg	tyr	val	45
	ser	his	phe	glu	pro	210	val	gly	ser	gly	gly	ala	tyr	
	arg	gly	pro	220	ser	thr	ala	trp	phe	arg	phe	gln	leu	50
	met	230	asp	asp	gln	asp	ala	arg	ala	thr	phe	tyr	240	gly
	ala	gln	cys	ser	leu	cys	thr	ser	leu		250	leu	trp	55
	ser	val	gly	arg	arg	gly	leu							60

Mutant or variant strains of *Pseudomonas putida* ATCC 53522 may be obtained by environmental selection pressure techniques, by UV irradiation, or by the use of mutagenic chemicals.

They may also be produced by genetic manipulation techniques, for example by the transfer of plasmid DNA to a multicopy host or by the excision of the chromosomal genes coding for the lipase from the cells of a lipase producing bacteria, followed by the cloning of said genes into a suitable vector molecule. The present invention encompasses such mutant, variant or cloned strains with retained, altered or enhanced ability to produce the lipase.

Thus, and as further described in Example 9, Figure 1 is a map of the 4.3 kb EcoRI fragment of pSNE4. The crosshatched box represents the signal peptide codons (codons -22 to +1), and the stippled region indicates the coding region for the mature Lipase 1 polypeptide codons +1 to +258. The postulated disulfide bonds are shown. The scale is in base pairs (bp). The region sequenced (an SphI fragment of 1363 bp) is indicated with a double arrow. The ATG initiation codon and TAA stop codon are also marked.

The cloning and expression of Lipase 1 illustrated by Example 9 is described herein since a preferred means for carrying out this invention (that is, producing the Lipase 1) is by cloning, and surprisingly high yields have been obtained by following the Example 9 procedure.

Lipase 1 has excellent hydrolytic activity and produces peracid from the substrate in the presence of a peroxide source, despite the hostile, oxidizing environment. It produces peracid even in the presence of anionic surfactants, which typically inhibit the activity of enzymes. Further, Lipase 1 has a higher ratio of peracid/acid than does a commercially available enzyme such as Lipase CES.

Lipase 1 may be obtained and used as a crude preparation from the fermentation of P. putida, but preferably is separated from other proteins and purified by means known to the art, such as by ion exchange and gel permeation chromatography, to yield substantially enzymatically pure Lipase 1. This is primarily because the crude fermentation broth of P. putida was found to include another enzyme (hereinafter "Lipase 2") in addition to Lipase 1.

While the crude preparation may be utilized in bleaches in accordance with the present invention, it is preferred to utilize a substantially pure Lipase 1 preparation. The two enzymes, Lipase 1 and Lipase 2, may be separated by means known to the art such as chromatography. They can be distinguished by their different hydrolysis rates for p-nitrophenyl butyrate and p-nitrophenyl caprylate. Lipase 1 may also be produced by cloning to express this enzyme through a host organism, such as E. coli, followed by octyl sepharose chromatography of the cloned Lipase 1, as is more particularly described hereinafter.

Lipase 2 is also novel, hydrolyzes glyceride substrates, and may be used in applications such as in fats and oils processing as a digestive aid.

The use of emulsifiers or surfactants is generally desirable as in other peracid bleach products. The use of emulsifiers is believed to be of particular value in establishing and maintaining a phase interface promoting interaction between the enzyme and glyceride substrate. Emulsifiers or surfactants can similarly be of value in establishing and maintaining the enzyme and substrate within the aqueous phase.

Anionic surfactants (generally also present in commercially available detergents) may be employed. Examples of such anionic surfactants include ammonium, substituted ammonium (for example, mono-, di-, and triethanolammonium), alkali metal and alkaline earth metal salts of C₆-C₁₈ fatty acids and resin acids, linear and branched alkyl benzene sulfonates, alkyl sulfates, alkyl ether sulfates, alkane sulfonates, olefin sulfonates, hydroxyalkane sulfonates, acyl sarcosinates and acyl N-methyltaurides.

Nonionic surfactants are also suitable for use within the enzyme perhydrolysis system of the invention. Nonionic surfactants include linear ethoxylated alcohols, such as those sold by Shell Chemical Company under the brand name NEODOL. Other nonionic surfactants include various linear ethoxylated alcohols with an average length of from about 6 to 16 carbon atoms and averaging about 2 to 20 moles of ethylene oxide per mole of alcohol; linear and branched, primary and secondary ethoxylated, propoxylated alcohols with an average length of about 6 to 16 carbon atoms and averaging 0 to 10 moles of ethylene oxide and about 1 to 10 moles of propylene oxide per mole of alcohol; linear and branched alkylphenoxy (polyethoxy) alcohols, otherwise known as ethoxylated alkylphenols with an average chain length of 8 to 16 carbon atoms and averaging 1.5 to 30 moles of ethylene oxide per mole of alcohol; and mixtures thereof.

Additional nonionic surfactants include certain block copolymers of propylene oxide and ethylene oxide, block polymers propylene oxide and ethylene oxide with propoxylated ethylene diamine, and semi-polar nonionic surfactants such as amine oxides, phosphine oxides, sulfoxides, and their ethoxylated derivatives.

Suitable cationic surfactants include the quarternary ammonium compounds in which typically one of the groups linked to the nitrogen atom is a C₈-C₁₈ alkyl group and the other three groups are short chained alkyl groups which may bear inert substituents such as phenol groups.

Further, suitable amphoteric and zwitterionic surfactants, which may contain an anionic water-solubilizing group, a cationic group and a hydrophobic organic group, include amino carboxylic acids and their salts, amino dicarboxylic acids and their salts, alkybetaines, alkyl aminopropylbetaines, sulfobetaines, alkyl imidazolium derivatives, certain quarternary ammonium compounds, certain quarternary ammonium compounds and certain tertiary sulfonium compounds. Other examples of potentially suitable zwitterionic surfactants can be found in Jones, U.S. Patent 4,005,029, at columns 11-15, which are incorporated herein by reference.

Other exemplary emulsifiers include water soluble or dispersible polymers, such as polyvinyl alcohol (PVA), polyvinylpyrrolidone (PVP), methylhydroxypropylcellulose (MHPC), etc. as well as bile and other natural emulsifiers.

Additional adjuncts of a wide variety may be considered for use in combination with the enzymatic

perhydrolysis system of the present invention, depending upon the specific application contemplated. For example, the enzymatic perhydrolysis system may be employed or included within a wide variety of cleanser applications or formulations such as straight bleach products, prewash products (which are often in liquid form) and even various hard surface cleansers.

For liquid formulations, it may be convenient to keep the hydrogen peroxide source separate from either the substrate or the enzyme, and preferably, from both. This could be accomplished by using a multiple chambered dispenser, such as that disclosed in U.S. Patent 4,585,150, issued April 29, 1986, to Beacham et al., and commonly assigned to The Clorox Company.

Suitable adjuncts may include fragrances, dyes, builders, stabilizers, buffers, etc. Stabilizers may be included to achieve a number of purposes. For example, the stabilizers may be directed toward establishing and maintaining effectiveness of the enzyme for original formation components or even intermediate products existing after the formulation is placed in an aqueous solution. Since enzymes may be hindered in hydrolysis of the substrates because of heavy metals, organic compounds, etc., for example, suitable stabilizers which are generally known in the prior art may be employed to counter such effects and achieve maximum effectiveness of the enzymes within the formulations.

The preferred pH level for aqueous solutions in which the enzymatic perhydrolysis system is dissolved is about 8-11, more preferably about 9-10. Buffering agents can be utilized in the invention to maintain the desired alkaline pH level for the aqueous solutions. Buffering agents generally include all such materials which are well known to those skilled in the detergent art. In particular, buffering agents contemplated for use in the present invention include, but are not limited to, carbonates, phosphates, silicates, borates and hydroxides.

The following experimental methods, materials and results are described for purposes of illustrating the present invention.

Example 1

(A) Seeding and Fermenting

A seed medium was prepared with 0.6% nutrient broth (Difco) and 1% glucose (pH 6.5). 100 ml of this medium was sterilized in 500 ml fernbach flasks. The flasks were each seeded with a loopful from an overnight culture of *P. putida* ATCC 53552 grown on nutrient agar, and placed on a Newbrunswick shaker at 250 rpm, 37° C for 12 hours. The incubated 12-hour culture was then seeded at appropriate volumes (1-10% v/v) into a 1 liter fermenter (250 ml working volume), a 15 liter Biolafitte fermenter (12 liters working volume), or a 100 liter Biolafitte fermenter provided with a temperature controller, RPM, airflow and pressure controller. The fermenter medium contained 0.6% nutrient broth (Difco), 0.3% apple cutin, and 0.2% yeast extract (Difco), with an initial pH of 6.5. The medium was adjusted to pH 6.8 and sterilized for 40 minutes before seeding. Bacterial growth and enzyme production were allowed to continue in the fermenter for 12-15 hours.

(B) Enzyme Recovery by Microfiltration

The crude fermentation culture was first filtered in a Amicon unit outfitted with two Romicon microporous membranes (0.22u) to remove cells. Remaining enzyme in the retentate which was bound to the cutin particles was removed by centrifugation. Total recovery approached 90%.

(C) Concentration and Dialysis of Whole Cell Filtrate

The recovered filtrate from the Amicon unit was concentrated to a volume of 3 liters on an Amicon ultrafiltration unit with two Romicon Pm 10 modules. The concentrated material was then dialysed with 20 liters of 0.01M phosphate buffer, pH 7.5, to remove salts and color. Recovery at this stage averaged about 80%. Total activity for this crude preparation was 8.68×10^6 units. A unit of lipase activity is defined as the amount of enzyme which results in an increase of absorbance at 415 nm of 1.0/minute when incubated at 25° C with 2.0 mM p-nitrophenylbutyrate in 0.1 M pH 8.0 Tris-HCl buffer containing 0.1 wt. % Triton X-100.

Example 2

Lipase Activity After Ultrafiltration and Diafiltration

The binding of three p-nitrophenyl substrates and the turnover kinetics were studied for the crude preparation of Example 1(C), where reaction conditions were 0.1M Tris with 0.1 wt. % Triton X-100, pH 8.0, at 25° C. The substrates were p-nitrophenyl caprylate, p-nitrophenyl laurate, and p-nitrophenyl palmitate, and the data is set out in Table 1.

Table 1

	Substrate	K_m (μ M)	V_{max} (μ mole/min/mg protein)
5	PNPC	214	802
	PNPL	167	214
10	PNPP	183	112

15 The Example 1(C) preparation was used in a variety of experiments, further described below, which showed utility with the enzymatic perhydrolysis system of the invention; however, the Example 1(C) preparation includes two enzymes designated "Lipase 1" and "Lipase 2". Lipase 1 is the better perhydrolase, and particularly preferred embodiments of the invention utilize substantially enzymatically pure preparations of Lipase 1. A separation and purification of the crude Example 1(C) preparation is described in Example 3, a complete separation of Lipase 1 and Lipase 2 is described in Example 4 (preferred to obtain substantially enzymatically pure Lipase 1), and an extremely pure sample of Lipase 1 preparation (i.e. analytically pure for sequencing) is described in Example 5.

Example 3

25 Partial Purification of Lipase 1 and Lipase 2 by Ion Exchange and Gel Permeation Chromatography

Lipase 1 was initially partially purified from *Pseudomonas putida* fermentation broth by DEAE Sephacryl chromatography followed by Sephadex G-100 gel permeation chromatography. A DEAE column was equilibrated in 10 mM sodium phosphate buffer, pH 8, and the crude protein was applied to the column in the same buffer. PNB (p-nitrophenyl butyrate) hydrolase activity that was not bound to the column was associated with Lipase 1. Lipase 1 thus obtained from the DEAE step was subjected to chromatography on Sephadex G-100 in 10 mM sodium phosphate buffer pH 8. Lipase 1 eluted from this column as a discrete peak, and was identified by PNB hydrolase activity as well as perhydrolytic activity.

35 Example 4

Complete Separation of Lipase 1 and Lipase 2 by Hydrophobic Chromatography

Lipase 1 may be separated completely from Lipase 2 by chromatography on hydrophobic resins. The enzyme solution of Example 1(C) after ultrafiltration and diafiltration was adjusted to 0.5M NaCl and applied to a 0.8 x 7 cm octyl Sepharose column equilibrated in 10mM Tris(Cl) pH 8, 0.5M NaCl and washed to remove unbound protein. The following washes were then employed: 10mM Tris(Cl), pH 8, 2M urea; 10mM Na phosphate pH 8; 10mM phosphate, pH 8, 0.5M NaCl. After washing, the column was then developed with a linear gradient to 50% n-propanol. The column fractions were then assayed for activity on p-nitrophenyl butyrate (PNB) and p-nitrophenyl caprylate (PNC) in order to locate the lipase activities. Two lipases were clearly resolved, fraction 32 with a PNB/PNC ratio of 4.6 and fraction 51 with a PNB/PNC ratio of 1.40. These have been designated Lipase 1 and Lipase 2, respectively.

The fractions from this column were further analyzed by SDS gel electrophoresis. This analysis revealed that the two lipase activities track with 30,000 molecular weight bands characteristic of procaryotic lipases; in addition, Lipase 2 migrated as a doublet, and was clearly resolved from the single band of Lipase 1. Prior to sequence analysis, these two partially purified enzymes were separated from the high and low molecular weight contaminants by reverse phase chromatography.

Example 5

55 Purification of Lipase 1 by HPLC in Preparation for Enzyme Peptide Fragmentation

Prior to sequence analysis, the partially purified material of Example 3 was further purified by chromatography on a 4.8 x 100 mm, SynChromPak C4 reverse phase HPLC column. The system was equilibrated in 0.05% triethylamine (TEA) and 0.05% trifluoroacetic acid (TFA) (Solvent A) at 0.5 mL/min. 100 μ g to 1 mg of Lipase 1 was injected onto the column and the protein eluted at 0.5 ml/min with a compound gradient of Solvent A and n-propanol containing 0.05% TEA and 0.05% TFA (Solvent B) B/minute. A typical gradient was +5% from 0 to 20%B and then +0.5% B/minute to 60% B. All lipase is inactivated by this HPLC solvent system. The protein peaks eluting at about 35% Solvent B (Lipase 1) or at about 39% Solvent B (Lipase 2) were collected and used for further sequence analysis and preparation of CNBr fragments.

Example 6Preparation and Purification of Cyanogen Bromide Peptide Fragments for Amino Acid Analysis

The cyanogen bromide peptide fragments for amino acid sequence analysis were prepared and purified as follows. An aliquot of pooled Lipase 1 of Example 5 was dried in a SpeedVac centrifuge and then resuspended to 10 mg/ml in 8 M urea, 88% formic acid. The solution was mixed with one volume of 200 mg/ml CNBr in formic acid and incubated in the dark at room temperature for 2 hours. The product was then desalted into 40% solvent B:50% solvent A (see above) on a 0.8 x 7 cm IBF-TrisAcryl GF05 (coarse) column prior to reverse phase analysis. The peptides were initially separated using the same protocol as listed above for the purification of Lipase 1 by reverse phase. Solvent B, however, was changed to 35% propanol:65% acetonitrile (containing TEA and TFA). The initial digest and the peaks after chromatography were also analyzed on SDS/urea/pyridine gels followed by silver staining.

Two peaks were chosen from the chromatogram and subjected to rechromatography employing the conditions dictated above, this time on a 0.48 x 25 cm SynChromPak C4 column. After rechromatography, the purified peptides were held for sequence analysis.

Example 7Distinction of Lipase 1 from Lipase 2: Preparation of Cyanogen Bromide Fragments of Lipase 1 and Lipase 2

The purified fractions of Lipase 1 and Lipase 2 from the octyl Sepharose column (as in Example 4) were each diluted with 3 volumes of solvent A (0.05% triethylamine and 0.05% trifluoroacetic acid) and chromatographed (as in Example 5). As described in Example 4, the purified proteins were analyzed by SDS gel electrophoresis, and then pooled individually for comparison of the CNBr fragments and the N-terminal amino acid sequences of Lipase 1 and Lipase 2.

Example 8Specified Activity of Lipase 1

The specific activity of Lipase 1 was determined using the enzyme purified as in Example 4. Substantially enzymatically pure Lipase 1 has a specific enzyme activity of 3750 units per mg protein as defined in Example 1(C).

Example 9Preparation of Cloned Lipase 1 in E. ColiCloning of the Lipase 1 Gene of Pseudomonas Putida

The *Pseudomonas putida* strain (ATCC 53552) was grown overnight at 37°C in 200 ml LB (Luria Broth) medium. Cells were harvested by centrifugation and high molecular weight total DNA was prepared exactly according to a standard procedure as outlined by Birnboim et al., *Nucleic Acids Res.* 7, pp. 1513-1523 (1979). The DNA was digested to completion with *EcoRI* and ligated with T4 DNA ligase to a preparation of plasmid pBR322 (ATCC 37017) digested with *EcoRI* and dephosphorylated with bacterial alkaline phosphatase. All enzymes used for the manipulation of DNA were used according to the manufacturers' directions (New England Biolabs or Bethesda Research Laboratories). The ligated DNA was used to transform *E. coli* 294 (ATCC 31445) and ampicillin resistant (Ampr) colonies were selected. Accordingly, approximately 2×10^4 transformants were obtained (approximately 5×10^3 /plate). Plates were flooded with a solution of 4-methylumbelliferylbutyrate (10mM in 50 mM Tris-HCl, pH 8.0) and then illuminated with an ultraviolet lamp (wavelength 340 nm). Colonies which hydrolyzed the substrate to release the highly fluorogenic compound 4-methylumbelliferone appeared as intensely blue. Using this method 13 positive colonies were obtained. From each of these positive colonies a plasmid miniprep in Birnboim, *supra*. Each plasmid was digested with *EcoRI* and resulting fragments were resolved by polyacrylamide gel electrophoresis as described by Maniatis et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring, New York (1982). Most plasmids contained a single inserted fragment of 4.3 kb. The others contained other fragments in addition to this fragment. This result suggested that all positive colonies arose as a result of the expression of a common cloned gene contained on the 4.3 kb fragment. One of the plasmids which contained only the 4.3 kb fragment, designated pSNE4, was selected for detailed analysis.

Plasmid pSNE4 was digested with a variety of restriction enzymes which have 6 bp recognition sequences. These enzymes were used singly or in pairs. Analysis of the fragment sizes resulting from these experiments allowed the generation of a preliminary restriction endonuclease cleavage map of the 4.3 kb *EcoRI* insert of pSNE 4. This map is shown in Figure 1.

Several subfragments of the *EcoRI* insert of plasmid pSNE 4 which were at least 840 bp were subcloned into pBR322 in order to see if any contained a functional gene. Among the plasmids which were found to contain functional lipase genes was pSNES1, which contains a 2.3 kb *EcoRI/SalI* fragment from the *EcoRI* insert of pSNE4. (See Figure 1 for map location of this fragment).

The inserted fragment of pSNES1 was digested with further restriction enzymes and the resulting small

fragments were subcloned into bacteriophage M13 vectors, described by Roberts, Nucleic Acids Res., 12, supplement r167-r204 (1984), for sequencing by the dideoxy chain termination method of Sanger et al., Proc. Natl. Acad. Sci. USA 74, pp. 5463-5467 (1977). The sequence of the 1.36 kb of DNA between the SphI sites (refer to Figure 1), when translated in all possible reading frames, revealed a large open reading frame which includes the NH₂-terminal amino acid sequencing (residues 94-105 and residues 173-190). The methionine at position -22 is believed to be the initiation codon because it begins the code for a highly hydrophobic region typical of signal peptides. This signal peptide is presumably cleaved off during the secretion process after the alanine at position -1. The open reading frame ends at position 259, indicating that the encoded mature protein has 258 residues.

Regulated Expression of *P. putida* Lipase 1 Gene in *E. coli*

In order to achieve the regulated expression of the *P. putida* lipase gene in *E. coli*, an XbaI site was first introduced just before the ATG initiation codon by site directed mutagenesis, Adelman et al., DNA 2, pp. 183-193 (1983) in bacteriophage M13, and the modified gene was subsequently cloned into an expression vector which contains the strong tacII promoter, deBoer et al., Proc. Natl. Acad. Sci. USA 80, p. 2125 (1983). This was done by first digesting pSNES1 with SphI.

The 2.4 kb SphI fragment containing the entire lipase coding sequence was isolated and ligated into the replicative form (RF) of M13mp19 at its SphI site and the mixture was used to transfect *E. coli* JM101 (ATCC 33876). Clear plaques were picked and the bacteriophage (template) DNA in which the SphI fragment was present in a counterclockwise orientation was prepared. A partially complementary single-stranded fragment of DNA consisting of 50 nucleotides was synthesized which contained an XbaI site immediately 5' of the Lipase 1 ATG initiation codon. This 50-mer complements the template DNA from the -27 nucleotide position (before the ATG initiation codon) to the -9 position and from the +1 (the A of the ATG) to the +20 position. Between the -9 and the +1 positions, however, the sequence 5'-AACCTTCG-3' of the native lipase promoter region was to be changed to 5'-TATCTAGAATT-3' of the tacII promoter. Mutagenesis was performed.

Three hundred plaques were screened by hybridization with a ³²P-labeled synthetic oligonucleotide (5'-ATGAGGTATCTAGAATTATG-3') which spans the area of change. An RF of a positively hybridizing clone was prepared and cleaved with XbaI and SphI. A 1 kb XbaI/SphI fragment containing the gene was isolated and ligated into a vector obtained by digesting pHGH907tacII, described by deBoer, supra, with XbaI and SphI and isolating a 4.1 kb XbaI/SphI fragment containing the tacII promoter, and the ampicillin resistance gene. JM101 cells were then transformed with ligation mixture. An ampicillin resistant colony (containing plasmid pSNtacII--see Figure 2) was selected.

To determine the levels of cloned Lipase 1 synthesized by *E. coli*, JM101/pSNtacII was grown in 20 ml LB medium supplement with 1mM isopropyl-B-D-thiogalactoside (IPTG) for 10h at 37°C. 294/pBR322 was used as a negative control. The cells were separated from the culture supernatant by centrifugation and then fractionated into periplasmic and membrane/cytoplasmic components, Koshland, supra. Each fraction was tested for activity by p-nitrophenylbutyrate hydrolysis. β -lactamase (periplasmic marker) and β -galactosidase (cytoplasmic marker) were also measured, Gray et al., Proc. Natl. Acad. Sci. USA 81, pp. 2645-2649 (1984), in order to confirm the effectiveness of the cell fractionation procedure. Most of the Lipase 1 activity (74%) was present in the culture supernatant. Most of the cell associated enzyme was found in the cell wash fraction (17% of the total) with smaller amounts present in the periplasmic (2%) and cytoplasm/membrane (7%) fractions. No Lipase 1 activity was present in any fractions of the 294/pBR322 negative control culture.

Broth from the fermentation of *E. coli* strain JM101 harboring the plasmid pSNtacII was adjusted to 0.5M NaCl and purified by octyl Sepharose substantially as described when *P. putida* is fermented (Example 4), except the propanol gradient was eliminated and elution was achieved with 20% acetonitrile in 10mM Na phosphate, pH 8, 0.5M NaCl. The isolated product (cloned from the gene expressing the enzyme) was analyzed by SDS gels and migrated identically to the Lipase 1 product isolated from the original *Pseudomonas putida* strain.

Example 10

Preparation of Cyanogen Bromide Fragments from Cloned Lipase 1

Cyanogen bromide fragments from cloned Lipase 1 were prepared as follows. The product from the octyl Sepharose purification of cloned product (Example 9) was diluted with 3 volumes of solvent A and purified on the short C4 HPLC column, as described for Lipase 1 and Lipase 2 isolated from *Pseudomonas putida*. The product was analyzed on SDS gel.

Example 11

Comparison of CNBr Fragments of Lipase 1 from *P. putida* and CNBr Fragments from the Cloned Lipase 1 in *E. coli*

CNBr fragments of Lipase 1 from *P. putida* and CNBr fragments from the cloned Lipase 1 in *E. coli* were compared. HPLC purified Lipase 1 and 2 from *Pseudomonas* and the cloned Lipase 1 were each hydrolyzed by CNBr as described in Example 6 above. The products were analyzed by SDS/urea/pyridine electrophoresis. The results indicate the cloned protein is clearly Lipase 1. Lipase 1 isolated from *P. putida* (as in Examples 4-5)

was shown to be identical to the cloned Lipase 1 isolated from *E. coli* by the following criteria: (a) Lipase 1 from either organism was isolated by the same chromatographic procedure (as in Example 4); (b) the amino acid sequences of the N-terminal of the Lipase 1 isolated from either organism were the same; (c) the CNBr fragment pattern showed that the Lipase 1 and Lipase 2 are clearly distinguished and that the CNBr fragments of Lipase 1 from either *P. putida* or *E. coli* are identical; (d) the p-nitrophenyl butyrate and p-nitrophenylcaprylate substrate activity ratio of Lipase 1 from both bacterial sources is the same; and (e) the hydrolysis/perhydrolysis ratio with tricaprylin as substrate is the same for Lipase 1 as isolated from both organisms.

Example 12

Assay for Peracid Generation

An assay was developed for determining peracid generation by monitoring the oxidation of o-phenylene diamine ("OPD"). The oxidation of OPD by peracid is much quicker than by H_2O_2 , and results in an absorbance increase at 458 nm. The assay procedure was to withdraw 0.1 ml of the reaction mixture being tested, to add 0.2 ml OPD solution (although in some instances, the OPD was added to the initial reaction mixture), incubate at room temperature for 5 minutes, add 0.9 ml $CHCl_3/CH_3OH$ (1/1 v/v), centrifuge 1 minute, and read the absorbance at 458 nm. The standard plot (for C_8 peracid) was linear up to at least 36 ppm peracid.

Example 13 illustrates that a "crude" preparation including the preferred enzyme ("Lipase 1") and the other, but less preferred enzyme ("Lipase 2"), has acceptable hydrolytic activity in the presence of either 30 ppm peroctanoic acid or up to 800 ppm hydrogen peroxide.

Example 13

Stability of Lipase in the Presence of Hydrogen Peroxide and Peracid

A quantity of the Example 1 enzyme preparation (including a mixture of Lipase 1 and Lipase 2, as in Example 1(C)), 1 mg/ml, was combined with 0.5 wt. % trioctanoin, 100mM NaCl, and 10mM sodium phosphate to form a 2 ml reaction volume with pH 10 for each of five samples. The reaction mixture for each sample was maintained at 30°C. The five samples were tested for hydrolytic activity as follows. One sample had 30 ppm of peroctanoic acid added and the hydrolytic activity of the enzyme was determined (μ mole NaOH/min), as was that of a control (with no peroctanoic acid added). Two other samples had hydrogen peroxide added (400 ppm and 800 ppm, respectively) and the hydrolytic activity determined (μ mole NaOH/min) as was that of a control. Table 2 sets out the data.

Table 2

Sample	Added	Hydrolytic Activity (μ mole NaOH/min)
1	(Control) 0 ppm peroctanoic acid	0.48
2	30 ppm peroctanoic acid	0.242
3	(Control) 0 ppm hydrogen peroxide	0.51
4	400 ppm H_2O_2	0.413
5	800 ppm H_2O_2	0.379

As illustrated by Table 2, the hydrolytic activity of this crude enzyme preparation was reduced by the presence of peroctanoic acid and by hydrogen peroxide; however, the enzyme preparation was considered to be sufficiently hydrolytically active despite the hostile, oxidizing environment.

In a similar manner, the same quantity of a commercially available enzyme (K-30) was tested by adding 400 ppm or 800 ppm hydrogen peroxide to reaction volumes of 2 ml, each with 0.5 wt. % trioctanoin, 100 mM NaCl, and 10 mM sodium phosphate. The hydrolytic activity was determined (μ mole NaOH/min), as was that of a control. Table 3 sets out the data for comparison with Table 2.

Table 3

	<u>Sample</u>	<u>Added</u>	<u>Hydrolytic Activity</u> <u>(μmole NaOH/min)</u>
5	6	(control) 0 ppm H_2O_2	0.223
10	7	400 ppm H_2O_2	0.135
	8	800 ppm H_2O_2	0.056

15 Comparison of the Tables 2 and 3 data shows that the inventive enzyme is considerably more stable (less sensitive) in the presence of an oxidizing environment than the previously known enzyme.

Example 14A illustrates that a substantially pure preparation of the preferred enzyme (Lipase 1) is not inactivated, and has excellent hydrolytic activity, in the presence of 400 ppm hydrogen peroxide and 4-7 ppm peroctanoic acid generated from the perhydrolysis of trioctanoin. Example 14B illustrates the effect of pH on the perhydrolysis to hydrolysis ratio.

Example 14A

Stability of Lipase 1

25 Another enzyme preparation as in Example 4 (substantially pure Lipase 1) was tested in both the presence and the absence of hydrogen peroxide. The hydrolysis rate was measured as μ moles/ml/10 min. Each sample had substrate of 1.0 wt. % sulfate ("SDS"), and o-phenylenediamine (OPD) at 4 mg/ml. The reaction volume for each sample was 2 ml, and the temperature was 27°C. The substantially pure Lipase 1 preparation did not experience any inactivation by the presence of hydrogen peroxide at 400 ppm or from the presence of peracid which was generated due to perhydrolysis.

Example 14B

Effect of pH

35 An enzyme preparation as in Example 4 was tested under the following constant reaction conditions: 1 wt. % trioctanoin in 0.1 wt. % SDS, 380 ppm H_2O_2 , 1 μ g/ml enzyme, 2 mg/mg OPD, with a reaction volume of 5 ml at 27°C. Each reaction volume had the pH adjusted to a value between 8 and 11, and the perhydrolysis and hydrolysis values determined as μ mole/ml/5 minutes. The optimum pH appears to be about pH 10.

Example 15 shows that the novel enzyme has greater enzymatic reactivity to a lipid than it does to an ester.

Example 15

Peracid Generation from Various Substrates

45 Each sample had 380 ppm H_2O_2 , OPD of 2 mg/ml, 1 μ g/ml enzyme (including a mixture of Lipase 1 and Lipase 2, prepared as in Example 1), 1 wt. % substrate (as an emulsion with SDS in a substrate/SDS ratio of 10:1) in 40 mM sodium phosphate buffer. The pH was 9.0 (pH stat), the temperature 27°C, and the reaction volume was 5 ml. Table 4 illustrates the data as μ mole/ml/10 min.

Table 4

	<u>Substrate</u>	<u>Hydrolysis</u> <u>(Acid</u> <u>Production)</u>	<u>Perhydrolysis</u> <u>(Peracid</u> <u>Production)</u>
55	trioctanoin	4.24	0.42
	methyl octanoate	0.66	0.11

60 As illustrated by the data, the novel enzyme has increased reactivity to the triglyceride substrate (with respect to the simple ester), and is shown to be a lipase.

Many commercially available enzymes are inhibited by the presence of anionic surfactant. Indeed, an anionic surfactant such as SDS is routinely used to solubilize proteins in procedures as SDS electrophoresis by binding to the protein molecules, converting them to rod-like shapes, and masking their native charge with the

SDS negative charge. Since many, if not most, commercially available detergents include anionic surfactants, the ability of an enzyme to maintain its hydrolytic activity despite the presence of an anionic surfactant is an important advantage.

Example 16 illustrates the retention of hydrolytic activity by an enzyme in accordance with the invention in the presence of anionic surfactant, and the inhibition of activity by a commercially available enzyme for comparison.

Example 16

Comparison of Activities of Lipase 1 and Lipase K-30 in the Presence of Anionic Surfactants

Samples were prepared having the same composition and/or reaction conditions except for the particular enzyme. Comparison samples were prepared with the commercially available enzyme Lipase K-30, said to be obtained from *Aspergillus niger*, available from Amano, at 8.7 µg/ml. The inventive enzyme preparation was as in Example 1 at 16.8 µg/ml (to approximate the hydrolysis rate of Lipase K-30). Each sample had trioctanoin as an emulsion with SDS in a weight ratio of 10:1. Sodium phosphate buffer (10 mM) was present, the pH was 10, the temperature was 25°C, and the reaction volume for each same was 2 ml. The data showing trioctanoin hydrolysis for each enzyme in the presence of SDS is set out in Table 5.

Table 5

Amounts of Substrate (wt. %)	Substrate Hydrolysis (µl 0.1N NaOH/min)
With Inventive Enzyme:	
0.01	2
0.05	7
0.1	11
0.5	13
1.0	12
Substrate Hydrolysis (µl 0.1N NaOH/min)	
Amounts of Substrate (wt. %)	
With Commercially Available Enzyme:	
0.01	1
0.05	7
0.1	7.5
0.5	3
1.0	1.5

As can be seen by the data of Table 5, the two different enzymes provide about comparable hydrolysis at 0.05 wt. % substrate, but the commercially available enzyme was inhibited by the increasing amount of SDS when the substrate amount increased beyond about 0.1 wt. % trioctanoin. That is, the perhydrolysis rate was relatively poor for Lipase K-30. By contrast, the inventive enzyme was substantially unaffected by the presence of the anionic surfactant.

In tests analogous to Example 16, but with emulsifier (polyvinyl alcohol), both the commercially available Lipase K-30 and the inventive enzyme exhibited good hydrolytic rates.

Example 17

Comparative Enzymatic Peracid Generation

Peracid generation was determined for an enzyme preparation as in Example 3, and compared with the peracid generation of two commercially available enzymes in the presence of 0.5 wt. % SDS. Each test sample had 480 ppm hydrogen peroxide, enzyme at 6 µg/ml, trioctanoin substrate at 5 wt. % and 0.5 wt. % SDS. The constant pH was 10 and the temperature was 25°C. Table 6 sets out the perhydrolysis profile of the inventive enzyme.

Table 6

	<u>time (min)</u>	<u>peracid generated (ppm)</u>
5	2	3.9
	4	7.2
	6	8.1
	8	9.0
10	10	9.9

By contrast, the amount of peracid generated with commercially available Lipase CES (said to be derived from *Pseudomonas fl.*, available from Amano) remained substantially constant and low (about 0.5 ppm peracid), while the amount of peracid with commercially available Lipase K remains substantially constant and even lower (about 0.3 ppm peracid).

The ratio of perhydrolysis to hydrolysis is very important. One wishes the highest possible conversion to peracid from substrate. The inventive enzyme has a higher ratio of peracid/acid than a commercially available enzyme such as Lipase CES. This means the inventive enzyme more efficiently utilizes its substrate for peracid generation and thus less of the substrate need be present in bleaching formulations.

Example 18Comparison of Perhydrolysis and Hydrolysis Activities of Lipase 1 and Currently Available Commercial Lipase

Perhydrolysis and hydrolysis were measured in samples containing 400 ppm hydrogen peroxide, 0.12 M HPO_4^{2-} , pH 10.0, and at various concentrations of either a commercially available lipase (CES from Amano) or the inventive enzyme (from Example 1(C) preparation). Perhydrolysis was measured by thiosulfate titration at 14 min. of reaction. Hydrolysis was measured simultaneously by continuous titration using the pH stat. The quantity of substrate was 12.5 wt. % trioctanoin with 0.75 wt. % PVA. The inventive enzyme provided more peracid for a lower substrate hydrolysis, which showed it more efficiently utilized the substrate than did the Lipase CES.

The crude preparation illustrated by Example 1(C) has been discovered to include two enzymes designated "Lipase 1" and "Lipase 2". The perhydrolysis to hydrolysis ratio of one with respect to the other for trioctanoin substrate is different, and Lipase 1 is the better perhydrolyase when compared to Lipase 2. Example 19 illustrates these ratios.

Example 19Effect of Surfactants on the Hydrolytic and Perhydrolytic Activities of Lipase 1 and Lipase 2

Four samples were prepared, and each contained 1 wt. % substrate, 0.1 wt. % surfactant (either SDS or PVA), 400 ppm H_2O_2 , and 4 mg/ml OPD. The reaction volumes were 2 ml, pH 9.0, and temperature 27° C. Either Lipase 1 (from Example 3 preparation or from Example 9 preparation) or Lipase 2 (from Example 4) was added to the samples. Table 7 sets out the data.

Table 7

	<u>Surfactant</u>	<u>Lipase 1 (p-nitrophenyl butyrate hydrolysis units)</u>	<u>Lipase 2 (p-nitrophenyl butyrate hydrolysis units)</u>	<u>Perhydrolysis to Hydrolysis Ratio</u>
	SDS	4	0	0.19
	PVA	4	0	0.14
55	SDS	0	20	0.010
	PVA	0	20	0.011
	SDS	12	0	0.12
60	PVA	12	0	0.076

As may be seen from the Table 7 data, Lipase 1 is a significantly better perhydrolase for trioctanoin substrate than Lipase 2 in the presence of the anionic surfactant.

Example 20 illustrates that excess of the novel enzyme appears to provide increased hydrolysis, but with no increase in peracid. Again, this shows an effective utilization of substrate.

Example 20

Effect of Enzyme Concentration Upon the Perhydrolysis/Hydrolysis Ratio

The substrate was 1 wt. % trioctanoin emulsified with 0.1 wt. % SDS, there was 400 ppm hydrogen peroxide, 4 mg/ml OPD, a pH of 9.0, a temperature of 25°C, and a reaction volume of 5 ml. Three different amounts of enzyme (prepared as in Example 4) were utilized, with the results illustrated in Table 8.

Table 8

Enzyme Level (PNB* units/ml)	Hydrolysis (umole/ml)	
	5 min	10 min
3	1.7	3.2
6	3.5	5.9
12	6.1	10.1
	Perhydrolysis (umole/ml)	
	5 min	10 min
3	0.42	0.54
6	0.51	0.78
12	0.57	0.87

*p-nitrophenyl butyrate hydrolysis

Thus, the perhydrolysis to hydrolysis ratios after 5 minutes were 0.25, 0.15 and 0.09, respectively, and after 10 minutes were 0.17, 0.13 and 0.09, respectively. The smaller amount of enzyme was thus more efficient.

When separated, Lipase 1 and Lipase 2 were found to have quite different hydrolysis rates (hydrolytic activity) for p-nitrophenyl butyrate and for p-nitrophenyl caprylate. Thus, the two novel enzymes can be distinguished by their ratios of p-nitrophenyl butyrate to p-nitrophenyl caprylate hydrolysis, as illustrated by Example 21.

Example 21

Hydrolysis Rates of Lipase 1 and Lipase 2 with p-Nitrophenyl Butyrate and p-Nitrophenyl Caprylate as Substrates

The reactions were performed in samples containing 0.1 M Tris HCl, pH 8.0 with 0.1 wt. % Triton X-100 nonionic surfactant (available from Rohm & Haas) at 25°C. The hydrolysis rates of 2.0 mM p-nitrophenyl butyrate (PNB) for Lipase 1 (as from Example 3), was 0.60 (Δ OD 415 nm/min.), while that of 2.0 mM p-nitrophenyl caprylate (PNC) was 0.09, for a PNB/PNC ratio of 7. By contrast, the hydrolysis rate of PNB for Lipase 2 at the same concentration was 0.54, of PNC at the same concentration was 0.44, for a PNB/PNC ratio of 1.

The novel enzyme has been shown to produce peracid in the presence of a broad range of surfactants even under hostile conditions for commercially available enzymes, such as the presence of anionic surfactants.

Example 22

Perhydrolysis Activity of Lipase 1 and With Two Commercially Available Lipases

Samples including either the novel enzyme (as in Example 1(C)), commercially available Lipase K or commercially available Lipase CES were dissolved in an aqueous solution containing a substrate (trioctanoin), hydrogen peroxide, and a mixture of surfactants (anionic and nonionic). The solutions were at room temperature and had a pH of 10.0. Perhydrolysis was calculated as ppm after 14 minutes, as set out in Table 9.

Table 9

Enzyme (l mg/ml)	Detergent* (% w/w)	Trioctanoin:Emulsifier (% w/w)	H ₂ O ₂ (ppm)	Perhydrolysis (ppm)
Novel enzyme	0.028	9.5:0	400	4.0
Novel enzyme	0.026	9.5:0.05 (sodium deoxycholate)	400	3.8
Novel enzyme	0.028	9.5:0.15 (sodium deoxycholate)	381	3.4
Novel enzyme	0.028	9.5:0.01 (sodium lauryl sulfonate)	397	3.3
Lipase K	0.028	9.5:0	505	0
Lipase CES	0.028	9.5:0	400	0
Lipase CES	0.026	9.5:6.9 (propylene glycol)	417	0

*45.1 wt. % CALSOFT F-90 (alkylbenzene sulfonate, available from Pilot Chemical Co.),
 40.8 wt. % SLS (sodium lauryl sulfate) and 14.1 wt. % NEODOL 25-7 (C₁₂-C₁₅ alcohol with an
 average ethoxylation of 7, available from Shell Chemical)

An enzyme in accordance with the invention is thus shown to provide strikingly better perhydrolysis in the presence of detergents including anionic surfactants.

The preceding examples are based on the use of a triglyceride substrate with the functional group (i) of the preferred substrates referred to above. Other glycerides included within that same functional group could be substituted for the triglyceride in the preceding examples. At the same time, additional substrates as defined above, particularly those included within the preferred functional groups of ethoxylated ester (ii) and propoxylated esters (iii) could be substituted for the triglyceride substrates in the preceding examples.

Further in connection with the preferred functional substrate groups (i), (ii) and (iii) referred to above, specific substrate examples within the first functional group are clearly apparent from the preceding examples.

Examples of substrates from the other two groups are demonstrated by the method of synthesis for a propoxylated ester set forth in Example 23.

Example 23

The procedure for preparation of a propylene glycol monoester of carboxylic acid includes the following steps:

(1) Salt Formation and Dehydration: One equivalent of carboxylic acid and 0.09 equivalents of sodium carbonate were combined in a round bottom flask equipped with a magnetic stir bar and an oil bath for heating. The slurry was heated under vacuum to 150°C with constant stirring for about one hour to achieve dehydration. The vacuum was released and the reaction cooled to room temperature.

(2) Esterification: The cooled acid/acid salt solution from step (1) was combined with about six equivalents of propylene oxide and heated on a warm oil bath at about 60°C under reflux for approximately eight hours to complete the esterification reaction. (Completion of the esterification reaction was confirmed by G.C. monitoring).

(3) The reflux condensate was removed and excess propylene oxide boiled off. About 200 milliliters of diethyl ether per 100 millimoles of acid were added and the resulting solution extracted in a separatory funnel with two volumes of 5% sodium carbonate. One volume of brine was then added. The ether layer was dried over sodium sulfate, filtered and rotary evaporated to produce the resulting ester product (typically about 90% pure).

Other examples of functionalized substrates according to functional substrate groups (ii) and (iii) can be produced by similar procedures.

Example 24 illustrates stain removal studies of several preferred formulations.

Example 24

Diagnostic evaluations of oxidant performance were performed with 100% cotton swatches stained with crystal violet as follows. Crystal violet (0.125 g) was added to 1.25 liters of distilled water. One hundred two-inch by two-inch undyed, 100% cotton swatches were added to the solution and agitated for eight hours. The cotton swatches (now dyed with crystal violet) were removed from the staining solution and rinsed repeatedly with cold tap water until the effluent was nearly clear. The stained swatches were then individually placed on aluminum foil, blotted with paper towels, and allowed to air dry.

Three preferred formulations of the invention were prepared, as were corresponding control compositions. The three inventive compositions and the three corresponding control compositions were each used to wash the stained cotton swatches and the stain removal performance evaluated for each. The performance results are summarized in Table 10.

Table 10

<u>Inventive Composition (a)</u>	<u>% Stain Removal</u>
0.06 wt. % trioctanoin	63.6
0.04 wt. % sodium dodecylsulfate	
100 ppm H ₂ O ₂	
1 µg/ml Lipase 1	
20 µM EDTA	
(pH = 10.5)	
<u>Control Composition (a)</u>	
0.06 wt. % trioctanoin	50.6
0.04 wt. % sodium dodecylsulfate	
100 ppm H ₂ O ₂	
20 µM EDTA	
(pH = 10.5)	
<u>Inventive Composition (b)</u>	
0.06 wt. % trioctanoin	80.4
0.04 wt. % sodium dodecylsulfate	
200 ppm H ₂ O ₂	
1 µg/ml Lipase 1	
20 µM EDTA	
(pH = 10.5)	

Control Composition (b)

0.06 wt. % trioctanoin	69.8	
0.04 wt. % sodium dodecylsulfate		5
200 ppm H_2O_2		
20 μ M EDTA		
(pH = 10.5)		10

Inventive Composition (c)

0.02 wt. % trioctanoin	67.4	15
0.02 wt. % sodium dodecylsulfate		
50 ppm H_2O_2		
5 μ g/ml Lipase 1		20
20 μ M EDTA		
(pH = 10.5)		25

Control Composition (c)

0.02 wt. % trioctanoin	52.2	
0.02 wt. % sodium dodecylsulfate		30
50 ppm H_2O_2		
20 μ M EDTA		
(pH = 10.5)		35

As may be seen from the data of Table 10, the inventive compositions provided improved stain removal benefits with respect to the control compositions even though the control compositions included the hydrogen peroxide component. These improved stain removals are due to the enzymatic perhydrolysis system and are particularly striking as occurring in the presence of anionic surfactant which inhibits many prior known commercially available enzymes.

Perhydrolysis or activated oxidant systems disclosed by various of the preceding examples can be used in combination with any of a wide variety of detergent formulations typically employed in cleaning fabrics. Thus, a typical heavy-duty built powdered detergent for U.S. laundry applications includes anionic and/or nonionic surfactants, phosphate or non-phosphate builders, buffering agents, and miscellaneous additives such as brighteners, perfume, proteases, and the like. The perhydrolysis system of the present invention may be used by itself or as a minor part of such a typical powdered detergent.

Typical heavy-duty built powdered detergents for European conditions have about the same nominal compositions as the U.S. counterparts, but product use concentrations are usually as 1.2% solutions in the washing machines (rather than the about 0.15% solutions typical in United States practice). A preferred formulation for the inventive perhydrolysis system packaged in combination with a typical detergent formulation is from about 3-30 wt. % source of peroxygen, about 0.6 to about 12 wt. % substrate, and about 0.001 to about 0.7 wt. % Lipase 1.

Under typical United States laundry use (where the detergent is usually free of oxidative bleaches and enzymes for stain removal), it is common practice to use a product containing oxidants (typically sodium perborate or sodium percarbonate) and enzymes (proteolytic and amylolytic) in addition to the detergent to improve performance. These oxidant-containing products are usually formulated to deliver about 25-50 ppm A.O. (hydrogen peroxide) at a product use concentration of about 0.17% solution in the washing machine. When the inventive perhydrolysis system is intended for use with detergent in wash water at temperatures from about 70°F to about 100°F, then a preferred formulation for the inventive perhydrolysis system preferably has the source of hydrogen peroxide, the substrate, and the Lipase 1 in a weight ratio between about 2400:200:1 and 48:20:1. A particularly preferred enzymatic perhydrolysis system of the invention has a weight ratio of sodium perborate tetrahydrate, trioctanoin and Lipase 1 in a weight ratio of 95:39:1. Such laundry

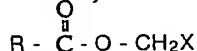
additive formulations generate about 25-50 ppm A.O. (hydrogen peroxide), 2-20 ppm A.O. (theoretical peracid) and 0.1 ppm to 10 ppm enzyme in the wash solution at a product use concentration of about 0.17%.

Claims

1. An enzymatic perhydrolysis system for in situ generation of peracid characterised in that it comprises:

- (a) an enzyme having hydrolase activity and being isolatable from Pseudomonas putida (ATCC 53552) or mutants and variants thereof;
- (b) a substrate being capable of hydrolysis by such an enzyme; and
- (c) a source of peroxygen which will react with (a) and (b) to produce peracid in the presence of a substrate-solubilizing aqueous solution.

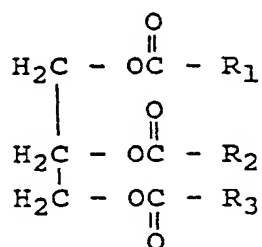
2. A system as claimed in claim 1 wherein the substrate has the structure:



wherein R is a substituent including at least one carbon atom and X is a functional moiety or a hydrocarbon group.

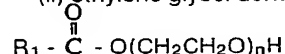
3. A system as claimed in claim 2 wherein X comprises a functionalized polyol or polyether; or X includes at least one carbon atom and at least one functional group.

4. A system as claimed in any of claims 1 to 3 wherein the substrate (b) is selected from :
(i) glycerides having the structure



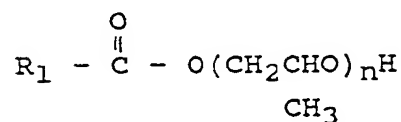
wherein R₁ is C₁-C₁₂, R₂ is C₁-C₁₂ or H and R₃ is C₁-C₁₂ or H;

(ii) ethylene glycol derivatives having the structure



wherein n = 1-10 and R₁ is defined as above; and

(iii) propylene glycol derivatives having the structure



wherein R₁ and n are defined as above.

5. A system as claimed in any of claims 1 to 4 wherein the substrate (b) is ordinarily incapable of substantial chemical perhydrolysis.

6. A system as claimed in claim 4 wherein R₁ is C₆-C₁₀, R₂ is C₆-C₁₀ or H and R₃ is C₆-C₁₀ or H.

7. A system as claimed in any of claims 1 to 6 wherein the substrate is normally insoluble in aqueous solution, and substrate-solubilizing aqueous solution includes an emulsifying agent.

8. A system as claimed in claim 7 wherein the emulsifier includes : a water soluble polymer; a cationic, nonionic, anionic, amphoteric or zwitterionic surfactant; bile salts; or mixtures of any of the foregoing.

9. A system as claimed in any of claims 1 to 8 wherein a buffer is also present, the buffer including a carbonate, phosphate, silicate, borate or hydroxide.

10. A system as claimed in any of claims 1 to 9 wherein the substrate (b) includes a triglyceride.

11. A system as claimed in any of claims 1 to 10 wherein the enzyme (a) is cloned from the gene expressing the enzyme, and has the amino acid sequence :

1	ala	pro	leu	pro	asp	thr	pro	gly	ala	10	pro	phe	pro	
								20						5
	ala	val	ala	asn	phe	asp	arg	ser	gly		pro	tyr	thr	
						30								
	thr	ser	ser	gln	ser	glu	gly	pro	ser		cys	arg	ile	
				40										10
	tyr	arg	pro	arg	asp	leu	gly	gln	gly		gly	val	arg	
		50												
	his	pro	val	ile	leu	trp	gly	asn	gly		thr	gly	60	
													ala	
														15
	gly	pro	ser	thr	tyr	ala	gly	leu	leu	70	ser	his	trp	
	ala	ser	his	gly	phe	val	val	80	ala	ala	ala	glu	thr	
														20
	ser	asn	ala	gly	thr	gly	arg	glu	met		leu	ala	cys	
						90								
	leu	asp	tyr	leu	val	arg	glu	asn	asp		thr	pro	tyr	
				100										25
	gly	110	thr	tyr	ser	gly	lys	leu	asn	thr	gly	arg	120	
													val	
	gly	thr	ser	gly	his	ser	gln	gly	gly	130	gly	gly	ser	
														30
	ile	met	ala	gly	gln	asp	thr	140	arg	val	arg	thr	thr	
	ala	pro	ile	gln	pro	150	thr	leu	gly		leu	gly	his	
						tyr								35
	asp	ser	ala	160	ser	gln	arg	arg	gln	gln	gly	pro	met	
	phe	170	leu	met	ser	gly	gly	gly	asp	thr	ile	ala	180	
													phe	40
	pro	tyr	leu	asn	ala	gln	pro	val	tyr	190	arg	arg	ala	
	asn	val	pro	val	phe	trp	gly	200	glu	arg	arg	tyr	val	
														45
	ser	his	phe	glu	pro	210	gly	ser	gly		gly	ala	tyr	
						val								
	arg	gly	pro	220	ser	thr	ala	trp	phe	arg	phe	gln	leu	
														50
	met	230	asp	asp	gln	asp	ala	arg	ala	thr	phe	tyr	240	
													gly	
	ala	gln	cys	ser	leu	cys	thr	ser	leu	250	leu	trp		
														55
	ser	val	gly	arg	arg	gly	leu							
														60

12. A system as claimed in any of claims 1 to 11 wherein the substrate (b) is trioctanoin or tridecanoin.

13. A system as claimed in any of claims 1 to 12 wherein the substrate (b) includes at least one glyceride

moiety, preferably being selected from diglycerides and triglycerides.

14. A process for bleaching a material characterised in that it comprises contacting the material with an aqueous solution and combining with the aqueous solution an enzymatic perhydrolysis system as claimed in any of claims 1 to 13.

15. A process as claimed in claim 14 wherein an emulsifier is present which is capable of at least maintaining or improving peracid yields achieved by the system in the absence of the emulsifier.

16. A process for the in situ generation of peracid characterised in that it comprises mixing a system as claimed in any of claims 1 to 13 with a substrate-solubilizing aqueous solution.

17. A process for the production of a system as claimed in any of claims 1 to 13 characterised in that it comprises mixing the constituents.

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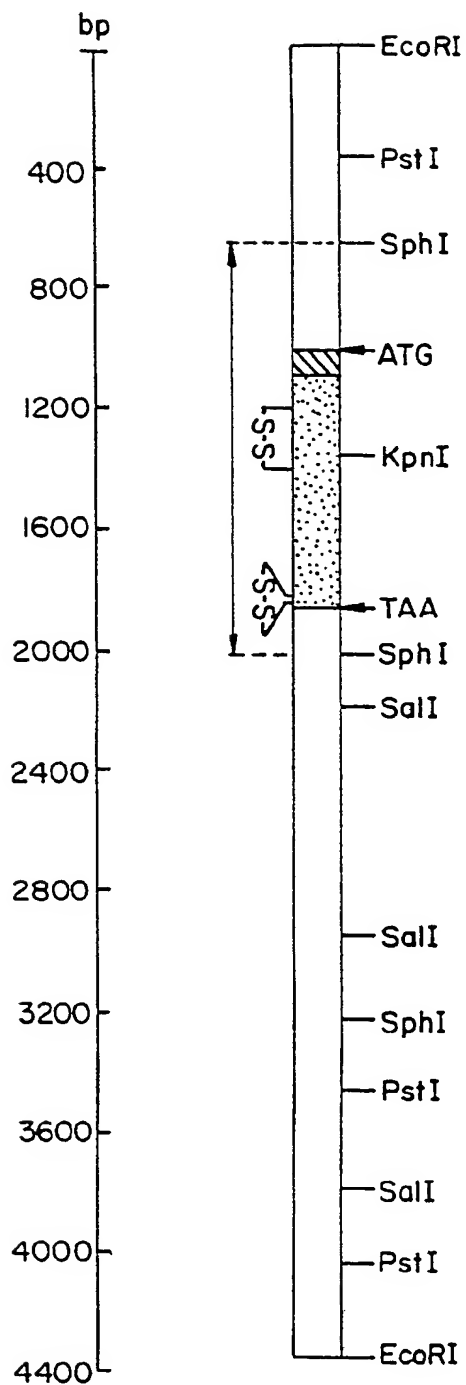


FIG. 1.

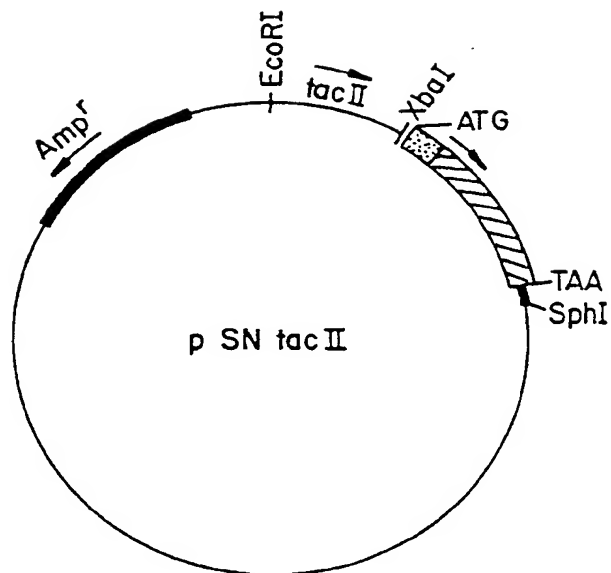


FIG. 2.



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Enzymatic peracid bleaching system.

An enzymatic perhydrolysis system, useful for bleaching, has a novel enzyme, a substrate, and a source of hydrogen peroxide, and provides in situ formation of peracid in aqueous solution. The substrate is selected for enzyme catalyzed reaction, and preferably is an acylglycerol with two or three fatty acid chains. The enzyme is hydrolytically and perhydrolytically active even in the presence of anionic surfactants, has lipase activity, and is isolatable from *Pseudomonas putida* ATCC 53552.

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EUROPEAN SEARCH REPORT

Application Number

EP 87 31 0130

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 4)
E	EP-A-0 253 487 (THE CLOROX CO.) * Whole document *	1-17	C 11 D 3/386 C 12 N 9/20
A, P	EP-A-0 212 976 (THE CLOROX CO.) * Whole document *		
A	S.N. LEWIS: "Peracid and peroxide oxidations", OXIDATION, vol. 1, editor R.L. AUGUSTINE, 1969, pages 213, 258, Marcel Dekker, Inc., New York, US * Whole chapter 5 *		
			TECHNICAL FIELDS SEARCHED (Int. Cl. 4)
			A 01 N C 11 D C 12 N
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 02-02-1989	Examiner CASTRO Y GONZALEZ R.F.
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